HAHM

International Journal Of Ayurvedic And Herbal Medicine 2:2 (2012) 366:379

Journal Homepage http://interscience.org.uk/index.php/ijahm

PROTECTIVE EFFICACY OF THE ETHANOLIC EXTRACT OF *ROSMARINUS OFFICINALIS* (Linn) AGAINST BRADYKININ INDUCED INFLAMMATION IN RATS

Kathiravan.S* and Shwetha.V.Kalava

Kongunadu Arts and Science College, Coimbatore, India

Correspondence Author :- Kathiravan subramanian Kongunadu Arts and Science College, Coimbatore, India

E-mail: kathitamizhanban@gmail.com.

The present study investigates the anti-inflammatory activity of ethanolic extracts of Rosmarinus officinalis Linn. The dried leaves of Rosmarinus officinalis were extracted with ethanol using soxhlet apparatus. The effect was observed on bradykinin induced paw edema in female SD rats. Inflammation was induced by subcutaneous injection of freshly prepared 0.1ml bradykinin (1mg/ml) in the right hind paw. The two treatment group rats received 250mg/kg b.wt and 500mg/kg b.wt respectively orally, one hour prior to bradykinin induction. The increase in the paw thickness in the rats after induction with bradykinin was significantly lowered on treatment with the plant extract. The ethanolic extract was found to improve the antioxidant status in the animals in a dose dependent manner. The effect of the extract was compared with the effect of the standard drug indomethacin administration. Histopathological analysis of the hindpaw tissue supports the protective effect of the ethanolic extract of R. officinalis.

INTRODUCTION

Inflammatory diseases including different types of rheumatic diseases are very common throughout the world. Inflammation is inherent to the pathogenesis of a variety of diseases. An inflammatory response implicates macrophages and neutrophils, which secrete a number of mediators (eicosinoids, oxidants, cytokine and lytic enzymes) responsible for the initiation, progression and persistence of the acute or chronic state of inflammation (Lefkowitz *et al.*, 1999). Currently, non-steroidal anti-inflammatory drugs (NSAIDs) supplemented with steroid hormone remains the major recommended strategy for its treatment (Scott *et al.*, 1998). While these drugs transiently suppress inflammation and ameliorate symptoms, they do not significantly improve the long-term disease outcome. Furthermore, long-term treatment with NSAIDs may result in serious side effects, such as gastrointestinal ulcergenicity and renal morbidity (Pincus *et al.*, 1992). Much attention has been directed towards the characterization of the antioxidant properties of the plant extracts, their fraction and identification of the constituents responsible for inflammation (Valentao et al., 2001; Haraguchi et al., 1996). The anti-inflammatory agents of present use exert their effect through a spectrum of different modes of action possessing well known side and toxic effects. It is therefore essential

Rosmarinus officinalis, a member of the family Lamiaceae is a flowering plant that grows in Mediterranean countries, Southern Europe and in the Littoral region through minor Asia wildly (Derwich et. al, 2011). Previous studies have shown that rosemary essential oil had antimicrobial, antioxidant, anti-carcinogenic, cognition improving and certain glucose level lowering properties, which make it useful as a natural animal feed additive. Many compounds have been isolated from rosemary, including flavones, diterpenes, steroids, and triterpenes (Omabola olunranti okoh, 2010).

MATERIALS AND METHODS

to introduce new medicinal plants to develop cheaper drugs.

2.1 Chemicals

Bradykinin for induction was purchased from Sigma Aldrich, Bangalore, India. All the chemicals used for the study were of Analytical grade.

2.2 Preparation of Plant extract

The leaves of *Rosmarinus officinalis* were bought from the local market and shade dried. The dried leaves was powdered and extracted with Ethanol, Chloroform, Petroleum ether, Ethyl acetate, Acetone and Benzene solvents using soxhlet apparatus. The extract obtained was rotary evaporated and the powder was preserved in an air tight container and stored at 4°C for further use.

2.3 Phytochemical screening of the extract

The phytochemicals were analysed with the dried extract according to the published standard methods.

The amount of phenols present was analyzed by Singleton and Rossi, 1965, flavonoids by Lamaison and Carnat, 1990, flavonols by Nakamura *et al.*, 2003 and tannins by Robert *et al.*, 1971 procedures.

2.4 Bradykinin induced acute inflammation model

There are various components to an inflammatory reaction that can contribute to the associated symptoms and tissue injury. Oedema formation, leukocyte infiltration and granuloma formation represent such components of inflammation (Mitchell and Cotran, 2000). Histamine, 5-hydroxytryptamine and bradykinin are the first detectable mediators in the early phase of inflammation (Di and Willoughby, 1971). Enzymes in a local area of injury are activated to free bradykinin. The main function of bradykinin is to increase the sensation of pain. A secondary function of bradykinin is to promote the production of histamine and is that of increasing blood flow into the involved area by dilation of arteries and increasing capillary vessel permeability.

Bradykinin (BD) induced paw edema model is a widely used model to evaluate the anti-inflammatory potency of plant extracts.

2.5 Experimental design

Female Sprague Dawley rats weighing approximately 180-200g obtained from small animal breeding station, Thrissur, Kerala, India were used for the study. The animals were maintained under standard conditions of humidity, temperature ($25 \pm 2^{\circ}$ C) and light (12h light/dark). The animals were divided into five groups of six animals each. Inflammation was induced by sub plantar injection of bradykinin (Mahat and Patil, 2007).

Group I: Control

Group II: Inflammation was induced by subcutaneous injection of freshly prepared 0.1ml BD in saline into the right hind paw.

Group III: Treated with 250mg/kg body weight of ethanolic extract of *R.officinalis* (ROEtOH) orally, 1 hour prior to BD induction

Group IV: Treated with 500mg/kg body weight of ethanolic extract of *R.officinalis* (ROEtOH) orally, 1 hour prior to BD induction Group V: Treated with 10mg/kg body weight of indomethacin orally, 1 hour prior to BD induction.

The edema was measured after 1, 2, 3 and 4 h. After 4h, the rats were killed by cervical dislocation and then the whole liver, spleen, hind paw were removed and washed with ice-cold saline. The tissues were homogenized using 0.1M Tris-HCl buffer (pH7.4) to give 10% homogenate.

2.6 Measurement of edema

Paw thickness was measured using Vernier caliper before and after BD challenge in each group. Increase in paw thickness was calculated using the formula Pt-Po, where Po is the initial paw thickness at time to and Pt is the thickness at time t (3h). Percent inhibition was calculated by the formula, (1-pt/pc)*100, where pt is the increase in paw thickness of treated and pc is that of BD induced control.

2.7 Biochemical Estimations

Serum Nitric oxide (NO) was determined by method described by Green *et al*, (1982). Sodium nitroprusside in aqueous solution, at physiological pH, spontaneously generate nitric oxide, which interacts with oxygen to produce nitrite ions that is estimated spectrophotometrically at 540nm. The detection of CRP is more

sensitive and reliable indicator of inflammatory process. This test is based on the immunological reaction between CRP as an antigen and latex particles as described by Tillet *et al*, (1930). The amount of protein present in the sample were estimated using standard method of Lowry *et al*, (1957). The activity of the enzyme, Superoxide dismutase (SOD) was assessed by the method of Das *et al.*,(2000). The method involves generation of superoxide radical of riboflavin and its detection by nitrite formation from hydroxylamine hydrochloride. The enzymic activity of Catalase (CAT) was determined by the method described by Sinha, (1972). The Glutathione(GSH) content of the hind paw tissue was determined by using Ellman's reagent as described by Moron *et al.*, (1979) Vitamin C is a very effective free radical scavenger. It is estimated spectrophotometrically at 520nm by method described by Omaye *et al* (1975) The extent of lipid peroxidation was measured through malondialdehyde reactivity with thiobarbituric acid in acidic condition to generate a pink coloured chromophore which were read at 535nm as described by Niehius and Samuelsson, 1968.

2.8 Histopathological analysis

Rats were sacrificed and the hindpaw tissues were collected and preserved in 10% formalin immediately after removal from animal.

2.9 Statistical analysis

All the values obtained from animals are expressed as mean \pm SD. Statistical comparision was done at significance level, P<0.05 using SPSS package version 10.0. One way ANOVA followed by post hoc analysis of LSD was performed.

3. RESULTS AND DISCUSSION

In Indian system of medicine, certain herbs are claimed to provide relief of pain and inflammation. The claimed therapeutic reputation has to be verified in a scientific manner. The present study was designed to investigate the anti-inflammatory effects of the medicnal plant, Rosemary (*Rosmarinus officinalis*). Studies have shown that phenolic compounds such as catechin and quercetin were very efficient in stabilising phospholipid bilayers against peroxidation induced by reactive oxygen species (ROS) (Gülçin *et al.*, 2010). Flavonoids are a class of phenolics that exhibit powerful antioxidant effects in biological systems, including free radical scavenging and metal ion sequestering, but their effectiveness greatly depends on particular chemical features. However, it is widely accepted that phenols have complex pro- and antioxidant effects in vitro, depending on their structure and the assay system used, and it is often hard to predict their actual action (Rice-Evans and Miller, 1996; Rice-Evans *et al.*, 1995).

The results of the qualitative and quantitative analysis of phytochemical screening are given in the table 1 and table 2 respectively.

Phytochemical			Ι	Solv	ent		
		Eth anol	Chloro form	Petroleu m ether	Ethyla cetate	Acet one	Benzen e
Alkaloid s	Dragendrof f test	+	+	+	+	+	+

Table 1 Phytochemical qualitative analysis of Rosmarinus officinalis

			1				
	Wagner's test	+	-	+	+	+	-
	Meyer's test	+	-	-	-	-	-
Flavo	noids	+	+	+	+	+	+
Sapo	nins	-	-	-	-	-	-
Carbohydrat es	Fehling's reaction	+	+	+	-	-	+
	Benedicts test	+	+	+	+	+	+
	Molisch's reaction	+	+	-	-	-	+
Protein	Millon's reaction	-	-	-	-	-	-
	Biuret test	+	-	-	-	+	+
Phenols	Ferric chloride test	+	+	+	+	+	+
	Lead acetate test	+	+	-	-	-	-
	Libermann reaction	+	+	-	-	-	+
Steroids	Libermann -Burchards	-	-	-	-	-	-
	Salkowski reaction	+	+	-	-	-	+
Glyco	Glycosides		+	-	-	-	-
Res	ins	+	+	-	-	-	+
Tannins	Ferric chloride	+	+	+	+	+	+

	Lead acetate	+	+	+	+	+	+
Thio	ls	-	-	-	-	-	-

In the qualitative analysis of phytochemicals, it was found that the ethanol extract gave better results and hence the ethanolic extract of *Rosmarinus officinalis* was chosen to carry out the further study.

Total p	Total phenols		noids	Flavonols	Tannins	Condensed
						Tannins
mg /g (GAE)	mg/g (CE)	mg /g (QE)	mg /g (RE)	mg/g (CAE)	mg/g (CAE)	mg/g (CAE)
61.34 ± 2.37	55.50 ± 3.12	6.22± 0.23	10.40 ± 0.61	43.3 ± 3.05	18.9 ± 1.21	1.53 ± 0.12

Table 2Phytochemical constituents in the ethanolic extract of Rosmarinus officinalis

The paw thickness of the experimental animals of different groups was analyzed upon induction with BD at a concentration of 1mg/ml. The initial paw thickness of all the experimental animals was measured. The paw thickness of group I animals that served as normal control, was found to be 0.58 ± 0.004 cm. The group II animals that served as BD control group, there was observed a significant raise in the paw thickness to 0.90 ± 0.014 cm. This increase could be due to the inflammation induced by BD. Upon treatment with the ethanolic extract of *R.officinalis* there was observed a significant reduction in the paw thickness in the group III and IV animals to 0.81 ± 0.005 cm and to 0.75 ± 0.007 cm respectively. The results obtained were similar to those evidenced in the group V animals that were treated with indomethacin. Thus it could be said that our extract was able to reduce inflammation in comparison to the standard drug. The values are shown in table 3.

Table 3 Effect of ethanolic extract of Rosmarinus officinalis on paw thickness of the control and

experimental animals

Groups	Initial paw thickness (cm)	Paw thickness after 4 h (cm)	Increase in paw thickness (cm)
Bradykinin Control	0.58 ± 0.004	0.90 ± 0.014	0.32 ± 0.002
ROEtOH (250 mg/kg b wt)+BD	0.57 ± 0.007	0.81 ± 0.005 ^a	0.24 ± 0.006
ROEtOH (500 mg/kg b wt)+BD	0.58 ± 0.006	0.75 ± 0.007^{a}	0.17 ± 0.008^{a}

Indomethacin(10mg/kg b wt)+ BD 0.58 ± 0.006 0.73 ± 0.007^{a} 0.15 ± 0.010^{a}

Nitric oxide (NO) is a potent pleiotropic inhibitor of physiological processes such as smooth muscle relaxation, neuronal signaling, inhibition of platelet aggregation and regulation of cell mediated toxicity. It is a diffusible free radical that plays many roles as an effector molecule in diverse biological systems including neuronal messenger, vasodilatation and antimicrobial and antitumor activities (Hagerman *et al.*, 1998)

Serum NO was found to increase significantly at p<0.05 from group I animals. After treatment with the plant extract, the serum NO decreased from group I animals. The 500mg/kg bwt dose of ROEtOH was found to be more effective when compared to 250mg/kg b.wt dose of ROEtOH.

Both nitric oxide (NO) and prostaglandin E_2 (PGE₂) are pleiotropic inflammatory mediators that are overproduced and involved in the pathogenesis of chronic inflammations and infections. NO is synthesized from L-arginine by nitric oxide synthase (NOS) (Kohno *et.al.*, 2008).CRP is a member of the class of acutephase reactants, as its levels rise dramatically during inflammatory processes occurring in the body. This increment is due to a rise in the plasma concentration of IL-6, which is produced predominantly by macrophages

NO along with superoxide (O_2) and the products of their interaction, also initiates a wide range of toxic oxidative reactions causing tissue injury (Hogg, 1998). Likewise, the neutrophils too produce oxidants and release granular constituents comprising of lytic enzymes performing important role in inflammatory injury (Yoshikawa and Naito, 2000). Inhibition in the release of these mediators is a potential strategy to control inflammation and is implicated in mechanism of action of a number of anti inflammatory drugs including the representative ones like dexamethasone (Bourke and Moynagh, 1999).

Thus by reducing the levels of NO in the serum, the extract reduces BD induced inflammation. The results of the study was found to be in accordance with Yoke Keong *et al* .(2011) who reported the protective effect of *Bixa orellana* leaves. The *Bixa orellana* extract was found to inhibit bradykinin-induced inflammation through suppression of nitric oxide production. The hydroalcoholic extract of *Coronopus didymus* was found to reduce NO levels in the serum of the rats that were induced with BD(Busnardo *et al*., 2010).

The CRP level in serum of induced group was very high when compared to group I and other groups. Induced group showed a hike to $25.75\pm0.23 \ \mu g/ml$. Compared to group III, extract administration to group IV has more effect in treating inflammation which reduced the CRP level to $9.66\pm0.11 \ \mu g/ml$. The values are shown on table 4.

Table 4 Effect of ethanolic extract of Rosmarinus officinalis on the levels of serum nitric oxide and

CRP and in control and experimental animals

Parameter s	Control	Bradykinin (1mg/ml)	ROEtOH (250mg/kg b wt) + BD	ROEtOH (500 mg/kg b wt) + BD	Indomethacin (10 mg/kg) + BD
Nitric	7.05 ± 0.21	10.20 ±	$8.28 \pm 0.21^{\ a b}$	7.25 ± 0.32^{b}	$7.08 \pm 0.31^{\text{ b}}$

Kathiravan.S International journal of ayurvedic & herbal medicine 2(2) April . 2012(366-379)

oxide	b	0.20 ^a			
CRP	5.76 ± 0.17	25.75 ± 0.23^{a}	$14.54\pm0.33^{\text{ ab}}$	9.66 ± 0.11^{b}	6.71 ± 0.28^{b}

Values are expressed as mean \pm SD (n=6)

Units: Nitric oxide - mg/dl; CRP-µg/ml

Group comparison and statistical significance at p<0.05: ^a: Group I vs. II, III, IV, V

^{b:} Group II vs. I, III, IV, V

The generation of reactive oxygen species (ROS) by phagocytic leukocytes (neutrophils, monocytes, macrophages, and eosinophils) is one of the most important hallmarks in the inflammatory process. The ROS are mediators of cellular injury and are involved in the onset of cellular damage during endotoxemia (Ginn-Pease and Whisler, 1998; Forman and Torres, 2001). ROS are involved in a variety of cellular stress mechanisms. Several lines of evidence indicate that the redox status of cells participates in modulating NK and B activation (D'Acquisto *et al.*, 2002). A number of reports have shown that a broad range of antioxidants abolish NF-_B activation (Bai *et al.*, 2005)

In accordance with the present state of scientific knowledge, there is enough evidence that the excessive production of free radicals in the organism, and the imbalance between generation of reactive oxygen species and antioxidant defenses is related to processes such as aging and several diseases (Kasapoglu and Ozben, 2001; Mattson *et al.*, 2001).

SOD may play an important role in protecting cells against ROS .SOD is the first enzyme of the anti-oxidant process (Beyer *et al.*, 1991). SOD catalyzes the breakdown of O_2^{--} and H_2O_2 , removes singlet oxygen as well as O_2^{--} , prevents formation of OH (Fridovich, *1973*), and has been implicated as an essential defense against the potential toxicity of oxygen.

The H_2O_2 formed by SOD and other processes is scavenged by catalase that catalyzes the dismutation of H_2O_2 into water and molecular oxygen. (Sumanth and Rana, 2006)

Enzymic antioxidants such as Superoxide dismutase (SOD), Catalase (CAT) play important roles in scavenging of reactive oxygen species. The activities of SOD and Catalase was analysed in spleen, thymus and hindpaw samples to check the antioxidant status in experimental animals.

The results are presented in Table 5. The activities of SOD and CAT were observed to be significantly lowered (p<0.05) in the BD control group animals as compared with group I normal control animals. The prior treatment with ROEtOH at 250mg/kg bwt and 500mg/kg bwt was found to significantly improve the antioxidant status in a dose dependent manner. The results were comparable with that of the standard drug, indomethacin. A decrease in the activity of SOD and CAT may be due to enormous production of free radicals. Akira *et al.*, (1988) reported that increase in the production of free radicals namely superoxide anions and hydroxy radicals could suppress the activity of SOD and CAT in inflammatory conditions. Superoxide anions are thought to be involved in inflammatory reactions since they are produced by phagocytic cells (Babior *et al.*, 1973). These cells are reported to produce hydroxy radical (Salin and McCord, 1975) and singlet oxygen (Allen *et al.*, 1972).

Table 5 Effect of ethanolic extract of *Rosmarinus officinalis* on the activities of enzymic antioxidants in spleen, thymus and hind paw of experimental animals

Parameter s	Control	Bradykinin (1mg/ml)	ROEtOH (250mg/kg b wt) + BD	ROEtOH (500 mg/kg b wt) + BD	Indomethacin (10 mg/kg) + BD
Spleen					
SOD	$4.12\pm0.03^{\text{ b}}$	$0.52\pm0.02^{\:a}$	1.82 ± 0.04^{ab}	3.38 ± 0.12^{b}	3.95 ± 0.23^{b}
CAT	11.31 ± 0.09^{b}	$3.96\pm0.05^{\ a}$	7.07 ± 0.06^{ab}	$10.19 \pm 0.05^{\ b}$	11.25 ± 0.05^{b}
Thymus					
SOD	$5.91\pm0.09^{\:b}$	$3.04\pm0.03^{\:a}$	$4.28\pm0.04^{\text{ b}}$	$5.50\pm0.03^{\ ab}$	5.65 ± 0.03^{ab}
CAT	$12.45 \pm 0.11^{\ b}$	4.13 ± 0.22^{a}	9.12 ± 0.56^{b}	$12.09\pm0.91^{\text{ b}}$	11.95 ± 0.58^{b}
Hind paw					
SOD	5.48 ± 0.41^{b}	3.97 ± 0.06^{a}	4.62 ± 0.26	5.20 ± 0.17^{b}	5.26 ± 0.15^{b}
CAT	8.15 ± 0.23^{b}	3.37 ± 0.28^{a}	5.85 ± 0.31	7.56 ± 0.52^{b}	7.61 ± 0.48^{b}

Values are expressed as mean \pm SD (n=6) Units:

SOD- inhibition of 50% nitrite formation/min; CAT-µmole of H₂O₂ consumed/min;

Group comparison and statistical significance at p<0.05: ^a: Group I vs. II, III, IV, V

^{b:} Group II vs. I, III, IV, V

GSH acts directly as a free radical scavenger by neutralizing OH⁻, restores damaged molecules by hydrogen donation, reduces peroxides, and maintains protein thiols in the reduced state (Sies, 1986).

Vitamin C is an outstanding antioxidant in biological systems and powerful reducing agent, also involved as cofactor in numerous metabolic processes. Vitamin C, a chain-breaking antioxidant, protects biological membranes against reactive oxygen species (ROS) (Frei, 1989). Ascorbic acid protects cells against oxidative damage to essential molecules. In addition vitamin C may reduce carcinogenesis through stimulation of the immune system.

The levels of Total Reduced Glutathione and Vitamin C were analysed in spleen, thymus and hindpaw samples. The decreased levels of GSH and Vit C in the group II animals , was found to be improved significantly (p<0.05) on treatment with ROEtOH at both the doses. The 500mg/kg bwt dose was found to be more effective than the 250mg/kg bwt dosage. The values are shown in table 6.

Table 6 Effect of ethanolic extract of *Rosmarinus officinalis* on GSH and Vitamin C in the spleen, thymus and hind paw of experimental animals

Kathiravan.S International journal of ayurvedic & herbal medicine 2(2) April . 2012(366-379)

Groups	Control	Bradykinin (1mg/ml)	ROEtOH (250mg/kg b wt)	ROEtOH (500 mg/kg b wt)	Indomethacin (10 mg/kg) + BD
			+ BD	+ BD	
Spleen					
GSH	1.81 ± 0.16^{b}	$1.43\pm0.05^{\:a}$	$1.64 \ 0.09^{ab}$	1.69 ± 0.06^{b}	1.72 ± 0.06^{b}
Vit C	$1.13\pm0.05^{\text{ b}}$	0.83 ± 0.04^{a}	0.95 ± 0.03^{ab}	$1.04\pm0.05^{\text{ b}}$	1.08 ± 0.02^{b}
Thymus					
GSH	2.11 ± 0.09^{b}	$1.22\pm0.05^{\:a}$	1.77 0.09 ^{ab}	$2.09\pm0.12^{\text{ b}}$	1.98 ± 0.08^{b}
Vit C	3.55 ± 0.05^{b}	$1.43 \pm 0.11^{\ a}$	2.90 ± 0.19^{ab}	3.33 ± 0.23^{b}	$3.28\pm0.21^{\text{ b}}$
Hind paw					
GSH	$1.14{\pm}0.10^{b}$	1.07 ± 0.0	$1.34\pm0.07^{\text{ ab}}$	1.43 ± 0.05^{ab}	$1.5\pm0.05^{\ ab}$
Vit C	$1.22\pm0.07^{\text{ b}}$	$0.73\pm0.04^{\text{ a}}$	$0.85\pm0.05~^{ab}$	$0.97\pm0.08^{\text{ ab}}$	$1.10\pm0.05^{\text{ ab}}$

Values are expressed as mean \pm SD (n=6) Units: GSH, Vitamin C - μ g/ml or mg protein;

Group comparison and statistical significance at p<0.05: ^a: Group I vs. II, III, IV, V

^{b:} Group II vs. I, III, IV, V

Lipid peroxidation can inactivate cellular components and plays an important role in oxidative stress.

Lipid peroxides are formed by auto-oxidation of polyunsaturated fatty acids found primarily in cell membranes. An increase in the level of lipid peroxides in tissues, therefore, reflects membrane damage (Kawamura *et al.*, 1992).

The levels of lipid peroxides in serum, spleen, thymus and hindpaw was found to be significantly (p<0.05) elevated in the BD induced group and the levels were restored in the treatment groups (Group III and IV) indicating the effect of the extract. The values are shown in table 7.

Table 7 Effect of ethanolic extract of *Rosmarinus officinalis* on lipid peroxide levels in serum, spleen, thymus and hindpaw of experimental animals

Groups	Control	Bradykin in (1mg/ml)	ROEtOH (250mg/kg b wt) + BD	ROEtOH (500 mg/kg b wt) + BD	Indomethacin (10 mg/kg) + BD
Serum					
LPO	8.35 ±	$18.39 \pm$	14.21 ± 1.35^{ab}	9.68 ± 0.65^{b}	8.84 ± 0.36^{b}

Kathiravan.S International journal of ayurvedic & herbal medicine 2(2) April . 2012(366-379)

	0.20^{b}	1.29 ^a			
Spleen					
LPO	${\begin{array}{*{20}c} 1.07 \pm \\ 0.06^{b} \end{array}}$	$\begin{array}{c} 1.82 \pm \\ 0.04^{a} \end{array}$	1.18 ± 0.04^{ab}	1.11 ± 0.05^{b}	1.10 ± 0.08^{b}
Thymus					
LPO	$\begin{array}{c} 1.07 \pm \\ 0.06^{b} \end{array}$	2.12 ± 0.04^{a}	1.58 ± 0.08^{ab}	1.09 ± 0.06^{b}	0.97 ± 0.08^{b}
Hind paw					
LPO	1.16 ± 0.03^{b}	$\begin{array}{c} 2.36 \pm \\ 0.10^a \end{array}$	$2.14\pm0.08^{\text{b}}$	$1.84 \pm 0.07^{\; b}$	$1.73\pm0.10^{\rm a}$

Values are expressed as mean \pm SD (n=6)

Units: LPO - nmoles of MDA formed/min/ml or mg protein; Group comparison and statistical significance at p<0.05: ^a: Group I vs. II, III, IV, V ^{b:} Group II vs. I, III, IV, V

The hind paw tissue of the experimental animals was analysed for histological changes. The results are presented in slide.

The various architectural changes in the tissue were depicted as follows.

Slide 1 : Shows the section of hind paw tissue of Group I animals. The architecture reveals no obvious abnoramilty

Slide 2 : Shows the section of hind paw tissue of Group II (BD induced) animals. The architecture indicates severe inflammation with infilteration of neutrophils

Slide 3 : Shows the section of hind paw tissue of Group III (BD + R. officinalis

250mg/kg b wt) .The architecture shows mild inflammation

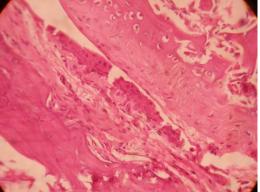
Slide 4 : Shows the section of hind paw tissue of Group IV (BD + R. officinalis

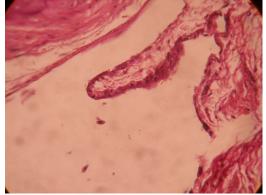
500mg/kg b wt) .The architecture indicates mild inflammation

Slide 5: Shows the section of hind paw tissue of Group V (BD + indomethacin 10mg/ kg b wt) . The architecture of the muscle fibres and synovium show no obvious abnormality.

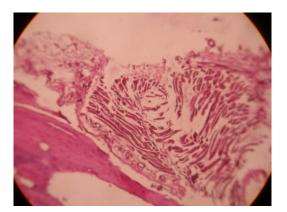
The histopathological investigation revealed the protective nature of the ethanolic extract of R officinalis against bradykinin induction.

Effect of the ethanolic extract of *Rosmarinus officinalis* on the histology of the hind paw tissue of experimental animals



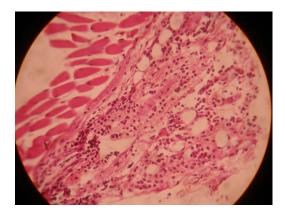


Slide 1: Normal Control. The architecture reveals no obvious abnormality Slide 2: Bradykinin induced The architecture reveals inflammation



Slide 3: Bradykinin induced + *R. officinalis* (250mg/kg b wt)

The architecture shows mild inflammation



Slide 4: Bradykinin induced + *R. officinalis* (500 mg/kg b wt)

The architecture shows no obvious abnormality



Slide 5: Bradykinin+ indomethacin (10mg/ kg b wt) The architecture of the muscle fibres and synovium show no obvious abnormality

Thus from the results of the biochemical analysis and histopathological investigation, it could be said that the ethanolic extract of *Rosmarinus officinalis* has potent anti-inflammatory action. This could be due to the various phytochemical constituents present.

ACKNOWLEDGEMENTS

The author thanks the Management of Kongunadu Arts and Science College, Coimbatore, India for the support and encouragement during the course of work.

REFERENCES

- Adel kadri, Zied Zarai, Ines Ben Chobba, Ahmed Bekir, Neji Gharsallah, Mohammed Damak *et al.* Chemical constituents and antioxidant properties of *Rosmarinus officinalis* L. essential oil cultivated from the south-western of Tunisia. Journal of Medicinal Plants Research 9 December, 2011 Vol. 5(29), pp. 6502-6508
- Akira Imadaya, Katsutoshi, Terasawa, Hiroyori, Tosa, Meguni, Okamots, Kazuo, Toriizuka. Erythrocyte antioxidant enzymes are reduced in patients with rheumatoid arthritis. J. Rheumatol., 1988; 15: 11.37.
- Allen R.C, Stjernholm R.L and Steele R.H. Evidence for the generation of an electronic excitation state(s) in human polymorphonuclear leukocytes and its participation in bactericidal activity. Biochem. Biophys. Res. Commun., 1972; 47: 679–684.
- Aziza kamal genana, Haiko hanse, arthur smania junior, simone machado de souza Rosemary(*Rosmarinus officinalis*)- a study of the composition, antioxidant and antimicrobial activities of extracts obtained with supercritical carbondioxide Ciênc. Tecnol. Aliment., Campinas, 2008; 28(2): 463-469.
- Babior, B.M., Kipens, R.S. and Curnutte, J.T. The production by leukocytes of superoxide dismutase, a potential bactericidal agent. J. Clin. Invest., 1973; 52: 741–744.
- Bai S.K, Lee S.J, Na H.J, Ha K.S, Han J.A, Lee H.S, Kwon Y.G, Chung C.K and Kim Y.M. Carotene inhibits inflammatory gene expression in lipopolysaccharide-stimulated macrophages by suppressing redox-based NF-B activation. Experimental and Molecular Medicine., 2005; 37: 23–334.
- Bourke E and Moynagh P.N. Antiinflammatory effects of glucocorticoids in brain cells independent of NF-kappa B. Journal of Immunology, 1999; 163: 2113–2119.
- Busnardo TC, Padoani C, Mora TC, Biavatti MW, Fröde TS, Bürger C *et al.* Anti-inflammatory evaluation of Coronopus didymus in the pleurisy and paw oedema models in mice. J Ethnopharmacol. 2010 Mar 24; 128(2):519-25.
- D'Acquisto F., May M.J and Ghosh S. Inhibition of nuclear factor kappa B (NF-B): an emerging theme in anti-inflammatory therapies. Molecular Interventions., 2002; 2: 22–35.
- DiRosa M, Giroud J.P. and Willoughby D.A. Studies on the mediators of the acute inflammatory response induced in rats in different sites by carrageenan and turpentine. J. Pathol. 1971; 104: 15-29.
- Elhoussine derwich, Zineb benziane, Rachida chabir and Rachid taouil. Invitro antibacterial activity and GC/MS analysis of the essential oil extract of leaves of rosmarinus officinalis grown in Morocco. Int J Pharm Pharm Sci, 2011; 3(3):89
- Elhoussine derwich, Zineb benziane, Rachida chabir, Rachid taouil. Invitro antibacterial activity and GC/MS analysis of the essential oil extract of leaves of *Rosmarinus officinalis* grown in morocco. Int J Pharm Pharm Sci, 2011:Vol 3, Issue 3, 89-95.
- Forman, H.J. and Torres, M. Redox signaling in macrophages. Molecular Aspects of Medicine., 2001; 22: 189–216.
- Frei B, England L and Ames B. Proc. Natl. Acad. Sci. USA, 1989; 86: 6377–6381.
- Fridovich I. Superoxide dismutase. Annu Rev Biochem., 1974; 44:147–159.
- Ginn-Pease M.E and Whisler R.L. Redox signals and NF-_B activation in T cells. Free Radical Biology and Medicine., 1998; 25: 346–361.
- Green H.C.C, Wagner D.A and Glogowski J. Analysis of nitrate, nitrite and (¹⁵N) nitrate in biological fluids. Anal. Biochem 1982;126: 131-138.
- Gulcin I, Sat I. G, Beydemir S, Elmastas M and Kufreviog lu O I. Comparison of antioxidant activity of clove (*Eugenia caryophylata* Thunb) buds and lavender (*Lavandula stoechas* L.) Food Chem, 2004; 87: 393–400.

- Hagerman AE, Riedl KM, Jones GA, Sovik KN, Ritchard NT and Hartzfeld PW. High molecular weight plant polyphenolics(tannins) as biological antioxidants. J Agric and Food Chem. 1998; 46: 1887-1892.
- Haraguchi H, Saitao T and Ishikawa H. Antiperoxidative components in thymus vulgaris. Planta Med 1996; 62: 217-221.
- Hogg N. Free radicals in disease. Seminar in Reproductive Endocrinology, 1998; 16: 241–248.
- Kasapoglu M. and Ozben T. Alterations of antioxidant enzymes and oxidative stress markers in aging. Exp. Gerontol., 2001; 36: 209–220.
- Kawamura T, Ohisa Y, Abe Y, Ishimori A, Shineha R. and Yokota, K. Plasma lipid peroxides in the operation of oesophageal cancer. Rinsho-Ayori, 1992; 40: 881–884 (in Japanese).
- Keizo kohno, Masaki miyake, Osamu sano, Mari tanaka-kataoka, Shigeto yamamoto, Satomi koyamiyata *et al.* Anti-inflammatory and Immunomodulatory Properties of 2-amino-3H-phenoxazin-3one.Biol.Pharm.Bull. 2008;31(10): 1938
- Lamaison J.L.C and Carnet A. Teneurs en principaux flavonoids des fleurs de Crataegeus monogyna Jacq et de Crataegeus laevigata (Poiret D. C) en function de la vegetation. Pharm. Acta. Helv., 1990;65: 315-320.
- Lefkowitz D.L, Gelderman M.P and fuhrmann S.R. Neutrophilic lysozyme-macrophage interactions perpetuate chronic inflammation associated with experimental arthritis. Clin Immunol 1999; 91: 145-155.
- Lowry O.H, Roseobrough N.J, Farr A.L and Randall R.J. Protein measurement with the Folin's phenol reagent. J. Biol. Chem. 1957; 193: 265-275.
- Mahat M.A and Patil B.M. Evaluation of anti-inflammatory activity of methanol extract of *Phyllanthus amarus* in experimental animal models. Indian J. Phar Sci. Jan Feb. 2007; 33-36.
- Mattson M.P, Duan W, Pedersen W.A, Culmsee C. Neurodegenerative disorders and ischemic brain diseases. Apoptosis 2001; 6: 69–81.
- Mitchell R.N, Cotran R.S. In:Robinsons Basic Pathology, Harcourt Pvt. Ltd; New Delhi, India, 2000; ed.7;33-42.
- Moron M.S, Defierre J.W and Mannervik B.. Levels of glutathione, glutathione reductase and glutathione-S-transferase activities in rat lung and liver. Biochem. Biophys. Acta. 1979; 582: 67-68.
- Nakamura Y, Tsuji S and Tonogai Y. Analysis of proanthocyanidins in grape seed extracts, health foods and grape seed oils. J.Health.Sci. 2003; 49:45-54.
- Nguyen Thi Dung, Vivek K.Bajpai, Jung In Yoon, Sun Chul Kang. Anti-inflammatory effects of essential oil isolated from the buds of Cleistocalyx operculatus(Roxb.) Merr and Perry. Food and chemical toxicology 2009; 47: 449-453.
- Niehius W.G. and Samuelsson D. Formation of malondialdehyde from phospholipid arachidonate during microsomal lipid peroxidation. Eur. J. Biochem. 1968;6: 126-130.
- Omabola oluranti okoh. Chemical transformations and phytochemical studies of bioactive components from extracts of *Rosmarinus officinalis* L.2010; 35.
- Omaye S.T, Turnball T.D and Sauberlich H.E. Selected methods for the determination of ascorbic acid in animal cells, tissues and fluids. Methods. Enzymol. 1979; 62: 1-11.
- Pincus, T., Marcum, S.B., Callahan, L.F. Long-term drug therapy for rheumatoid arthritis in seven rheumatology private practices. II. Second line drugs and prednisone. The Journal of Rheumatology 1992; 19: 1885–1894.

- Rice-Evans C.A, Miller J and Paganga G. Antioxidant properties of phenolic compounds. Trends Plant Sci., 1997; 2: 152–159.
- Rice-Evans C.A, Miller N.J and Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. Free Radic Biol Med1996; 20: 933–956.
- Robert E.B. Method for estimation of tannin in grain sorghum. Agro. J., 1971; 63: 511.
- Salin M.L, McCord J.M. Free radicals and inflammation: protection of phagocytosing leucocyte by SOD. J. Clin. invest. 1975; 56: 1319.
- Scott D.L, Shipley M, Dawson A, Edwards S, Symmons D.P, Woolf A.D. The clinical management of rheumatoid arthritis and osteoarthritis: strategies for improving clinical effectiveness. British Journal of Rheumatology 1998; 37: 546–554.
- Sies H. Biochemistry of oxidative stress. Angewandt Chemie International Edition in English, 1991; 25: 1058-1071.
- Singleton V.L and Rossi J.A. Colorimetry of total phenolics with phosphotungstic acid reagents. Am. J. Enol. Viticul. 1965; 16: 144-158.
- Sinha A. K. Colorimetric assay of catalase. Anal. Biochem1972; 47: 389-394
- Sumanth, M and Rana, A.C.. Invitro antioxidant activity of hydroalcoholic extracts of *Taxarum* officinale roots in rats Indian J Pharmacol., 2006; 38(1): 54-5.
- Tillet W.S, and Francis T, J.Exter, Med, 1930; 52: 561.
- Valentao P, Fernandes E, Canvalho E, Andrade P.B, Seabra R.M and Bastos M.L. Antioxidant activity of Hypericum androsaenium infusion scavenging effect on superoxide radical, hydroxyl radical and hypochlorous acid. Biol Pharm Bull 2002; 25: 1324-1327.
- Veiga JF, Zunino, Patitucci MP, Pinto AC and Calixto JB. The inhibition of paw oedema formation caused by the oil of Copaifera multijuga hane and its fractions. Pharm. Pharmcol. 2006; 2042-2158
- Yoke Keong Y, Arifah AK, Sukardi S, Roslida AH, Somchit MN, Zuraini A. Bixa orellana leaves extract inhibits bradykinin-induced inflammation through suppression of nitric oxide production. Med Princ Pract. Epub 2011 Jan 20.20(2):142-6.
- Yoshikawa T, Naito Y. The role of neutrophils and inflammation in gastric mucosal injury. Free Radic Res 2000; 33: 785–794.